

ELEMENTAL ANALYSIS ON SALIVA SAMPLES OF PATIENTS WITH PERI- IMPLANTITIS USING TXRF

João Marcos Fávaro Lopes

<https://orcid.org/0000-001-7096-169X>

Avacir Casanova Andrello

<https://orcid.org/0000-0001-6943-8973>

Ítalo José Vitorino Netto

<http://lattes.cnpq.br/6927784105567490>

All content in this magazine is licensed under a Creative Commons Attribution License. Attribution-Non-Commercial-Non-Derivatives 4.0 International (CC BY-NC-ND 4.0).



Abstract: The goal of this paper is to examine the composition of saliva samples collected from patients suffering from peri-implantitis and try to find a correlation between the detected elements using TXRF and the disease. We did so for four samples from different regions of the patient's mouth were collected from four subjects. We then added a gallium internal standard to all the samples and let it dry in an oven to perform the measurement using a S2 Picofox from Bruker, then the obtained spectra were analyzed using the Spectra 7 software. By performing the analysis, we noticed that the elements detected on every sample were: Phosphorus, Sulfur, Chlorine, Potassium, Calcium, Iron, Zinc, Bromide and Rubidium. Titanium was also detected on 72% of the samples. Our initial assumption was that the concentration of titanium would be greater in the regions near the compromised implants due to corrosion. We actually found that the titanium concentration was greater at the gingival sulcus of health implants. The results also showed that the concentration of iron and sulfur were greater at the gingival sulcus of compromised implants than anywhere else in the mouth. With these findings, we can confirm that the TXRF is a reliable technique to analyze elements from trace to macro-concentrations and can be used to monitor salivary levels of several elements, but so far, we couldn't find a relation between peri-implantitis and implant corrosion.

Keywords: TXRF, Peri-Implantitis, Saliva.

INTRODUCTION

Mankind has used dental implants to replace missing teeth since immemorial times (SAINI, 2015). As science and technology developed throughout history, so have the materials used to develop new kinds of dental implants. Nowadays, the most common material used to this task is titanium (MOMBELI, 2018). This

material is chosen due to its biocompatibility with the oral environment. Titanium implants are surrounded by a layer of titanium oxide TiO_2 that grants protection against corrosion to the implant (SAINI, 2015).

Even though the titanium oxide provide protection to the implants, this layer can be compromised due to some factors such as mechanical wear, pH, bacteria, chemicals and other contaminants (SAFIOTI, 2017; SOLER, 2020). More specifically, bacteria present in the mouth produce toxins that acidify the implant surrounds and can cause dissolution of the titanium oxide layer (MATHEW, 2012). Once the protective layer wears off, the implant becomes exposed to the buccal environment and is now prone to corrosion.

Such disruption of the titanium oxide layer and consequent implant corrosion is what defines the disease known as peri-implantitis (SAFIOTI, 2017). This is an inflammatory disease that affects the peri-implant mucosa and the jaw supporting bone where the implant is installed (LANG, 2011). The diagnostic is given by the BOP (bleeding on probing) exam and radiography of the jaw (SCHWARZ, 2018).

We found papers that chose to analyze the behavior of this disease by employing spectroscopy techniques over tissue samples collect from the gingival mucosa (MERCAN, 2013; HE, 2016) or submucosal plaque (SAFIOTI, 2017). Our aim was to make a similar analysis but using saliva samples instead.

Saliva is a biological fluid whose analysis can provide several information about the subject's health (COLON, 1997). Therefore, performing the elemental analysis on saliva samples collected from individuals with peri-implantitis could provide interesting information about the disease's behavior. 99% of saliva's composition is water, while the remaining 1% is composed by several others

organic and inorganic substances (POLETTTO, 2021).

Due to the reduced size of the samples and low concentration of elements to analyze, the technique used to perform the analysis was the Total Reflection X-Ray Fluorescence (TXRF). The TXRF is an inexpensive multi-element method suitable for trace-element analysis with a simple sample preparation (VAN GRIEKEN, 2001). This technique was previously applied to saliva analysis by Abraham et al. (ABRAHAM, 2014), Cleto et al. (CLETO, 2016), Borella et al. (BORELLA, 1994), Zahir et al. (ZAHIR, 2006) and Poletto et al. (POLETTTO, 2021) among others.

In this study, we hypothesized that the elemental content of samples collected from different regions of different people can reflect the region where each sample was collected, i.e., higher Ti concentrations could indicate that the samples originated from an implant sulcus. The objective of this study was to apply multi-elemental TXRF analysis to the saliva samples and further evaluate them by using Principal Component Analysis (PCA) to check if the samples present some kind of relation. This paper is based on J. Lopes' master's thesis: "Análise Elementar em Amostras de Saliva de Pacientes com Peri-Implantite por TXRF" (LOPES, 2022).

MATERIALS AND METHODS

SAMPLE COLLECTION

Four samples were collected from different mouth regions from four subjects. The relation between patients and region from where the samples were collected can be seen on Table 1. All patients agreed to take part on this study and the samples were collected during routine dental exams.

TXRF SPECTROMETER

The device used to perform the measurements was the S2 Picofox (Bruker Corporation, Berlin, Germany), located in the Applied Nuclear Physics Laboratory at the Londrina State University, Parana State, Brazil. The system is a transportable TXRF spectrometer that dispenses cryogenic refrigeration, therefore is useful not only in a laboratory but also for on-site analyses in the field. The device performs simultaneous and fast determination of all elements from sodium to uranium and can detect concentration limits as small as ppb.

This spectrometer is specially indicated for environmental analysis, quality control, material science, medicine and biotechnology. The voltage, the current and the maximum power that can be applied to the molybdenum

| Region | Patient | | | |
|-------------------------|---------|---|---|---|
| | 1 | 2 | 3 | 4 |
| Parotid gland duct | X | X | X | X |
| Healthy implant sulcus | X | X | | X |
| Implant interior | X | | | X |
| Peri-implantitis sulcus | X | X | X | X |
| Total saliva | | X | X | |
| Peri-implant mucositis | | | X | |

Table 1: Relation between patients and samples.

X-ray tube built within the device are 50 kV, 600 μ A and 30 W, respectively.

The Picofox system is designed for sample carriers that are 3 cm of diameter and 3 mm in height. Among the types of samples carriers indicated for the device, we chose to use acrylic discs since they are cheap, disposable and presents a small blank on each measurement.

Once the saliva was deposited on the discs, 10 μ L of an internal standard of gallium at the concentration of 5 mg/L were pipetted over each sample. After that, the samples were taken to an oven at 55°C for approximately 35 minutes to dry out and then be measured. Each sample was measured three times for a live time of 100 seconds. The precision detection varied for each element on each measurement and is reported as the lower limit of detection (LLD).

QUANTIFICATION AND DATA ANALYSIS

Quantification of the samples was performed with respect to the internal standard using the Spectra 7 software, developed specifically to use alongside the S2 Picofox system. To validate the built-in calibration of the software we used two reference material: NIST 1640 and NIST 1643e. Mean concentrations and standard deviations were calculated with MatLab software and bar graphs comparing the concentrations of each element detected on the samples were plotted using Microsoft Excel. Principal Component Analysis (PCA) was also performed using MatLab.

Due to the high divergence encountered for concentration values along the samples, we opted to carry out the PCA using the z-scores of each value instead of the actual concentrations. The equation used to find these scores is written below,

$$Z = \frac{|X - \bar{X}|}{\sigma}$$

where Z represents the standardized concentration, X is the concentration to be standardized, \bar{X} is the mean concentration for a given element across all samples and σ is the standard deviation for this distribution. The module had to be taken since the PCA requires positive integers.

RESULTS AND DISCUSSION

The elemental concentrations detected by the Picofox system were determined by comparison between the detected counts for each element peak in comparison with measured for the internal standard. A typical spectrum is shown in Figure 1.

Along the measurements, the detected elements were: Al, Si, **P**, **S**, **Cl**, **K**, **Ca**, Ti, V, Cr, Mn, **Fe**, Co, Ni, Cu, **Zn**, As, Se, **Br**, **Rb** and Sr. Those written in bold were detected on all sixteen samples. Titanium wasn't detected on any sample from one of the patients but was detected on every other sample. Therefore, we decided to consider its behavior in our analysis.

The concentrations measured for each element on each sample are displayed on Table 2 alongside its respective z-scores. These concentration results are also displayed on the bar plots from Figures 2 through 5.

By examining the table, one can notice that the concentrations values encountered diverge very much from one another, therefore the standardization is necessary to proper analysis the elements behavior. Beyond that, the bar charts were plotted displaying two scales to aid on the visualization.

There are three immediate results that can be taken from the bar plots:

- The Parotid gland samples presents the lowest concentration of elements for all four subjects;

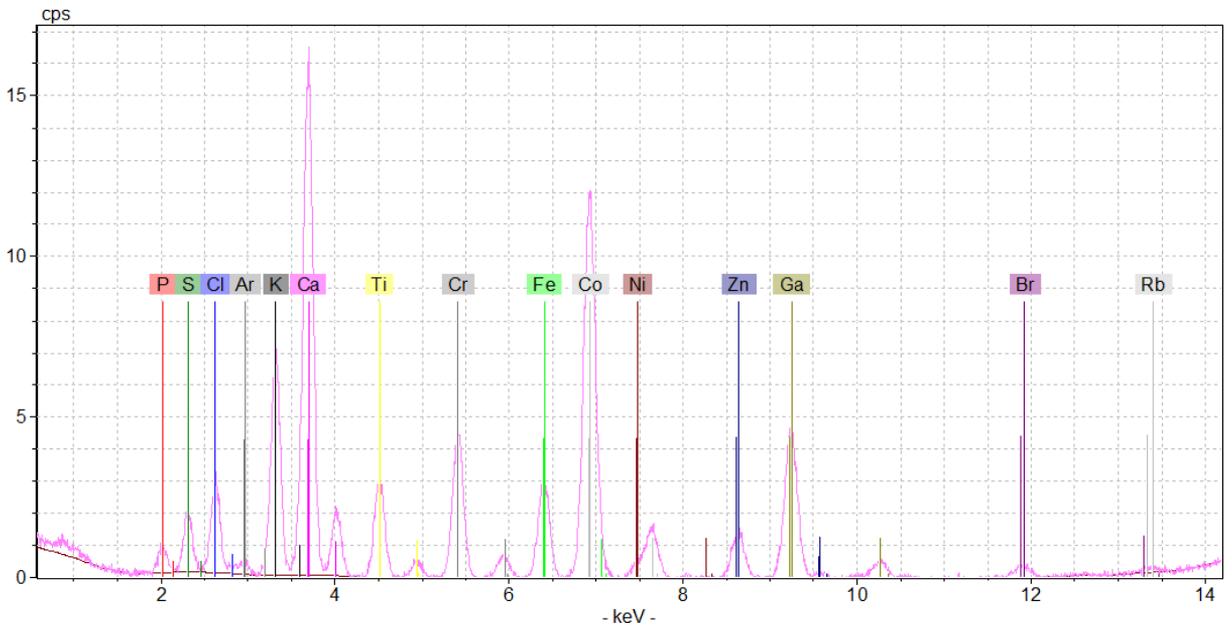


Figure 1: A typical TXRF spectrum for saliva samples.

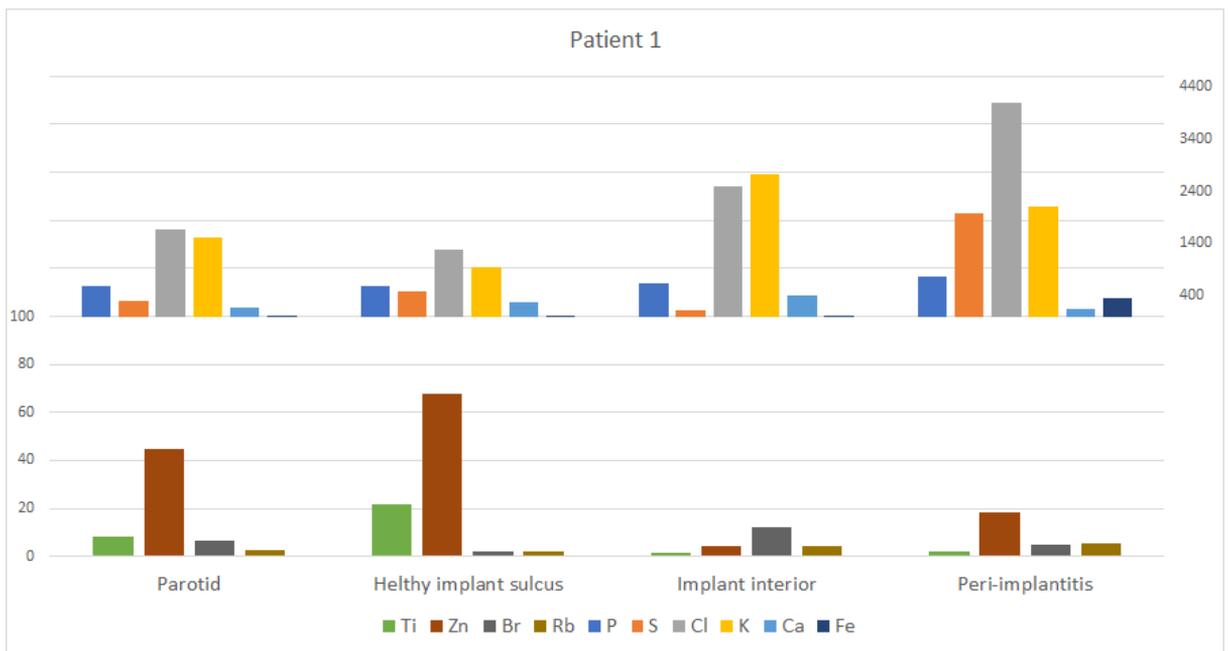


Figure 2: Measured concentrations for the first patient.

| | | | P | S | Cl | K | Ca | Ti | Fe | Zn | Br | Rb |
|-----------|-------------------------|---|----------|----------|----------|----------|---------|--------|---------|--------|--------|-------|
| Patient 1 | Parotid | C | 571,777 | 296,769 | 1657,553 | 1522,850 | 179,470 | 8,075 | 0,821 | 44,865 | 6,627 | 2,792 |
| | | Z | 0,314 | 0,172 | 0,381 | 0,167 | 0,062 | 0,428 | 0,326 | 1,792 | 0,265 | 0,344 |
| | Healthy implant sulcus | C | 575,615 | 488,519 | 1272,071 | 948,546 | 276,633 | 21,782 | 7,176 | 67,643 | 2,118 | 1,943 |
| | | Z | 0,325 | 0,177 | 0,046 | 0,325 | 0,712 | 2,179 | 0,253 | 2,990 | 0,668 | 0,114 |
| | Implant interior | C | 644,031 | 127,666 | 2494,580 | 2715,303 | 399,286 | 1,748 | 9,935 | 4,428 | 12,348 | 4,576 |
| | | Z | 0,515 | 0,479 | 1,107 | 1,190 | 1,532 | 0,380 | 0,221 | 0,336 | 1,450 | 1,306 |
| | Peri-implantitis sulcus | C | 766,180 | 1980,317 | 4105,117 | 2102,243 | 151,983 | 2,353 | 347,553 | 18,173 | 4,897 | 5,480 |
| | | Z | 0,855 | 2,886 | 2,505 | 0,664 | 0,122 | 0,303 | 3,659 | 0,387 | 0,093 | 1,793 |
| Patient 2 | Parotid | C | 276,844 | 16,208 | 56,593 | 873,042 | 66,253 | 0,153 | 0,215 | 0,624 | 4,456 | 1,003 |
| | | Z | 0,507 | 0,681 | 1,009 | 0,390 | 0,695 | 0,584 | 0,333 | 0,536 | 0,184 | 0,621 |
| | Peri-implantitis sulcus | C | 1316,917 | 1444,686 | 2165,709 | 3651,269 | 314,938 | 1,028 | 4,416 | 16,511 | 12,816 | 3,630 |
| | | Z | 2,388 | 1,913 | 0,822 | 1,993 | 0,968 | 0,472 | 0,285 | 0,300 | 1,547 | 0,796 |
| | Healthy implant sulcus | C | 799,671 | 387,188 | 1920,310 | 2240,240 | 210,808 | 0,930 | 2,796 | 3,926 | 10,311 | 2,944 |
| | | Z | 0,948 | 0,008 | 0,609 | 0,783 | 0,272 | 0,485 | 0,303 | 0,362 | 1,028 | 0,426 |
| | Total saliva | C | 608,426 | 171,543 | 2038,566 | 2978,192 | 535,077 | 0,940 | 2,683 | 6,113 | 10,775 | 4,886 |
| | | Z | 0,416 | 0,399 | 0,712 | 1,416 | 2,440 | 0,484 | 0,304 | 0,247 | 1,124 | 1,473 |
| Patient 3 | Peri-implantitis sulcus | C | 91,797 | 159,053 | 219,692 | 159,300 | 9,740 | 0,000 | 0,693 | 0,896 | 0,463 | 0,390 |
| | | Z | 1,022 | 0,422 | 0,867 | 1,002 | 1,073 | 0,604 | 0,327 | 0,522 | 1,011 | 0,951 |
| | Peri-implant mucositis | C | 27,036 | 14,705 | 4,363 | 49,371 | 5,948 | 0,000 | 0,000 | 0,314 | 0,142 | 0,026 |
| | | Z | 1,202 | 0,684 | 1,054 | 1,096 | 1,098 | 0,604 | 0,335 | 0,552 | 1,077 | 1,148 |
| | Total saliva | C | 299,723 | 120,379 | 345,758 | 873,741 | 56,053 | 0,000 | 0,347 | 0,815 | 6,117 | 1,021 |
| | | Z | 0,443 | 0,492 | 0,758 | 0,389 | 0,763 | 0,604 | 0,331 | 0,526 | 0,160 | 0,611 |
| | Parotid | C | 3,880 | 1,149 | 1,783 | 1,198 | 1,983 | 0,000 | 0,000 | 0,133 | 0,054 | 0,009 |
| | | Z | 1,266 | 0,709 | 1,056 | 1,138 | 1,125 | 0,604 | 0,335 | 0,562 | 1,096 | 1,157 |
| Patient 4 | Peri-implantitis sulcus | C | 245,275 | 515,837 | 1019,345 | 551,991 | 90,820 | 1,310 | 78,213 | 2,642 | 1,151 | 1,128 |
| | | Z | 0,595 | 0,226 | 0,173 | 0,665 | 0,531 | 0,436 | 0,564 | 0,430 | 0,868 | 0,553 |
| | Implant interior | C | 71,373 | 62,044 | 72,100 | 74,878 | 120,872 | 14,895 | 6,444 | 1,773 | 0,339 | 0,123 |
| | | Z | 1,078 | 0,598 | 0,995 | 1,074 | 0,330 | 1,299 | 0,261 | 0,476 | 1,037 | 1,095 |
| | Healthy implant sulcus | C | 280,696 | 382,042 | 725,989 | 421,222 | 175,575 | 22,395 | 4,854 | 3,414 | 1,296 | 0,859 |
| | | Z | 0,496 | 0,017 | 0,428 | 0,777 | 0,036 | 2,258 | 0,279 | 0,389 | 0,838 | 0,698 |
| | Parotid | C | 763,201 | 93,258 | 1400,752 | 2079,071 | 127,106 | 0,000 | 0,596 | 0,709 | 11,611 | 3,657 |
| | | Z | 0,847 | 0,541 | 0,158 | 0,644 | 0,288 | 0,604 | 0,328 | 0,532 | 1,297 | 0,810 |

Table 2: Concentrations and respective z-scores for each measurement.

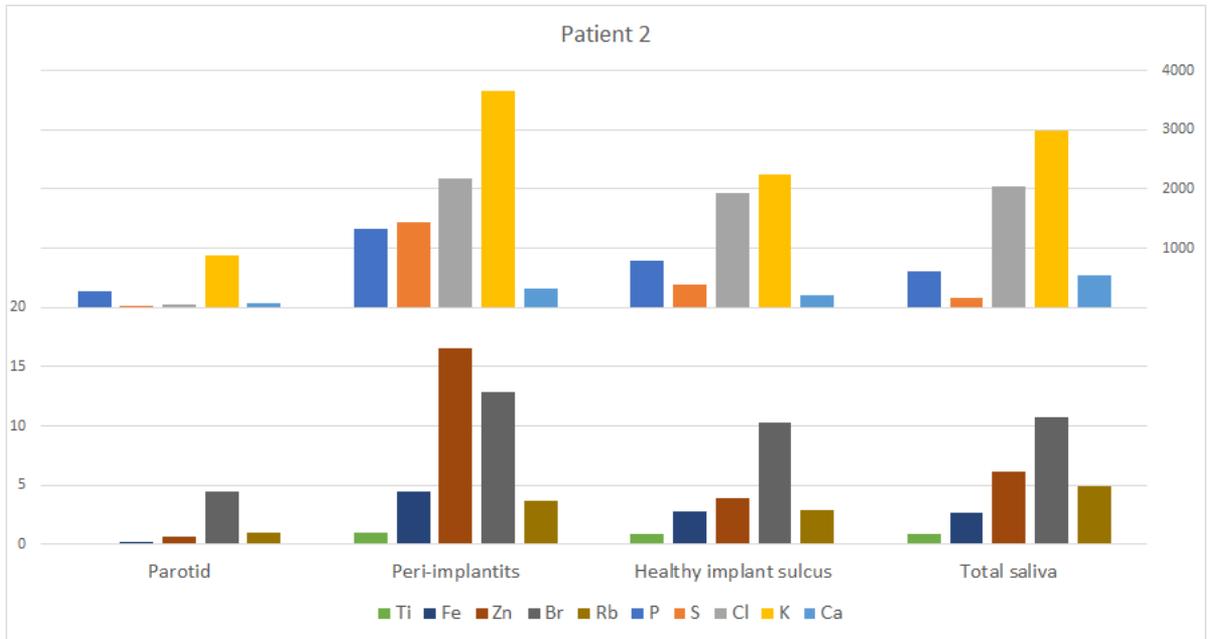


Figure 3: Measured concentrations for the second patient.

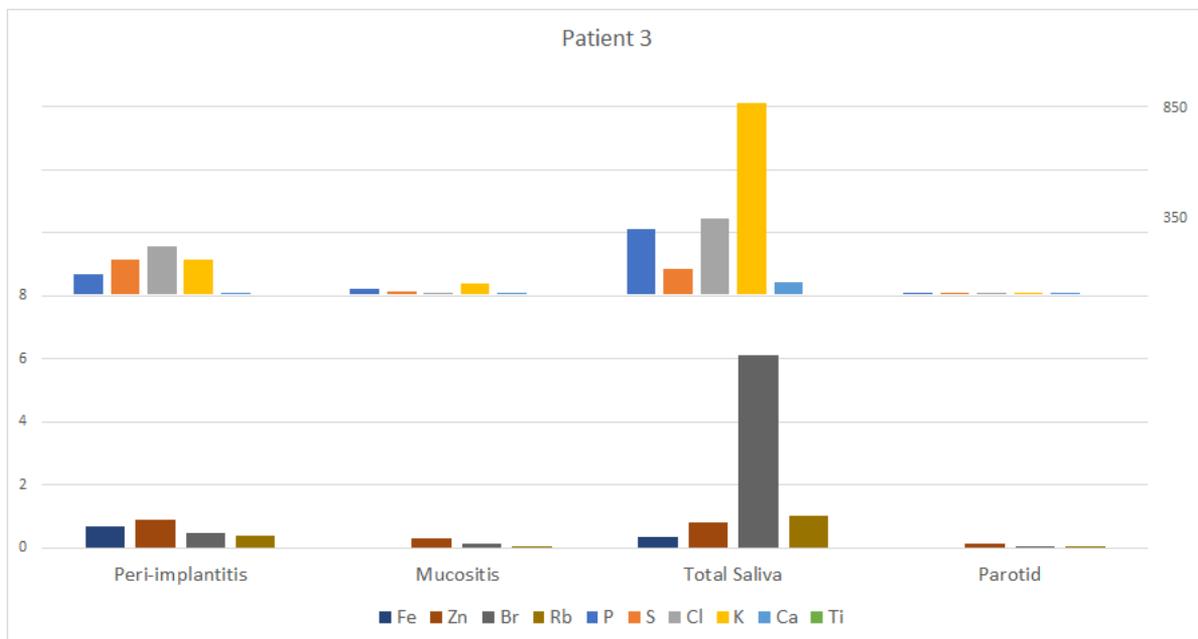


Figure 4: Measured concentrations for the third patient.

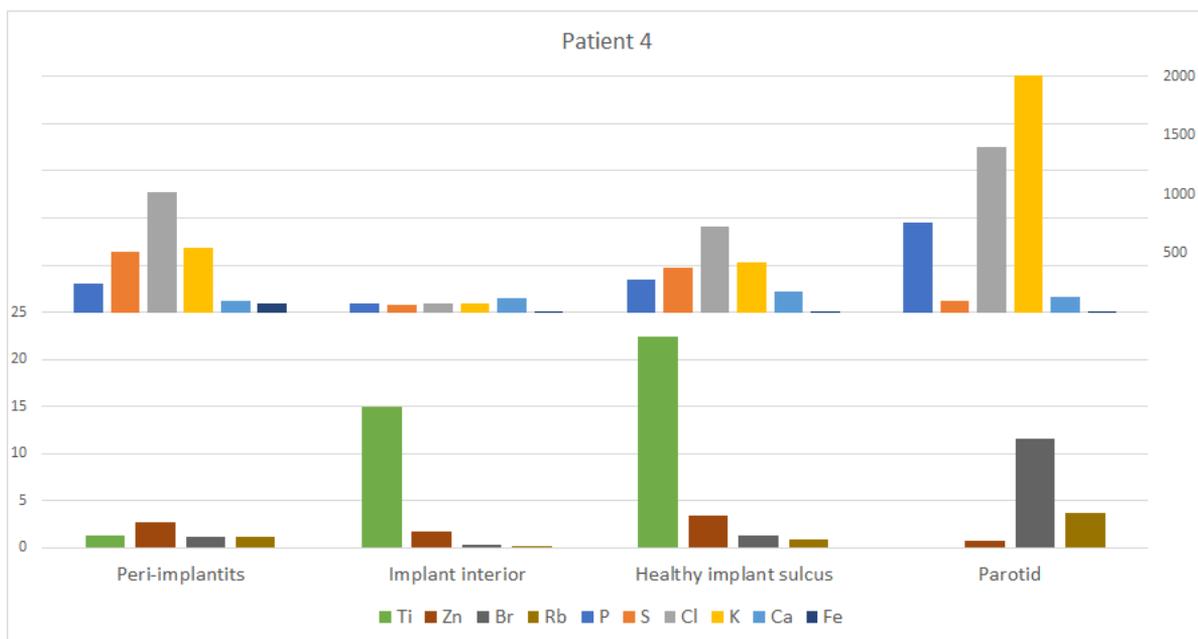


Figure 5: Measured concentrations for the fourth patient.

- The sulfur and iron concentration are higher on peri-implantitis sulcus than any other regions; and
- The titanium concentration is higher on healthy implant sulcus than on peri-implantitis sulcus.

The fact that the third result goes against what was found by Mercan et al. (MERCAN, 2013) suggests that the titanium corroded from implants is deposited in the gum instead of being diluted on saliva.

Using the values obtained by calculating the z-scores we also performed the PCA for all samples, the result is shown on Figure 6.

Unfortunately, no discerning behavior was obtained, it is not possible to infer the regions from which the samples were taken because there are no cluster formations.

Therefore, even though the TXRF is a reliable technique for elemental composition and saliva analysis, we can't seem to find a relationship between a sample composition and its origin by using chemometric analysis (PCA).

ACKNOWLEDGMENTS

We would like to thank CNPq and CAPES for funding and financial support.

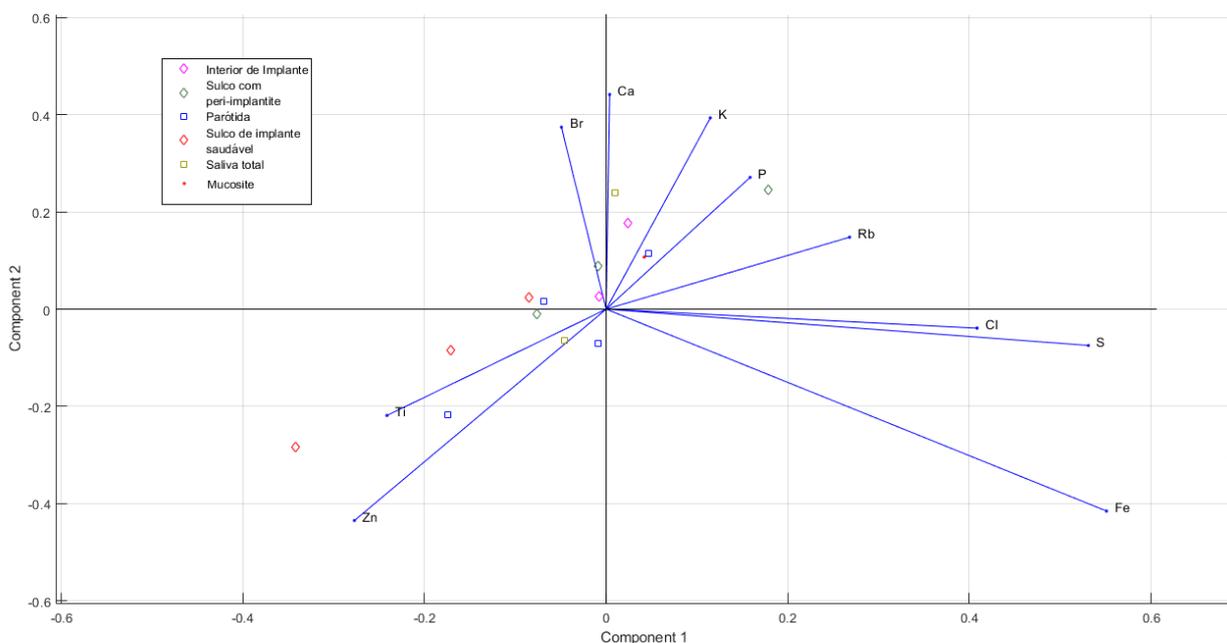


Figure 6: PCA for the standardized results.

REFERENCES

- Abraham, J. A., Sánchez, H. J., Grenón, M. S., & Pérez, C. A. (2014). **TXRF analysis of metals in oral fluids of patients with dental implants.** *X-Ray Spectrometry*, 43(4), 193-197.
- Borella, P., Fantuzzi, G., & Aggazzotti, G. (1994). **Trace elements in saliva and dental caries in young adults.** *Science of the total environment*, 153(3), 219-224.
- Cleto, D. A. M., Andrello, A. C., Netto, Í. J. V., & Appoloni, C. R. (2016). **Analysis of saliva and gingival crevice fluid by total reflection X-ray fluorescence (TXRF).** *X-Ray Spectrometry*, 45(4), 220-224.
- Colon, L. A. (1997). **Analysis of body fluids: urine, blood, saliva and tears.** In *Handbook of Capillary Electrophoresis Applications* (pp. 486-498). Springer, Dordrecht.
- He, X., Reichl, F. X., Wang, Y., Michalke, B., Milz, S., Yang, Y., ... & Högg, C. (2016). **Analysis of titanium and other metals in human jawbones with dental implants—A case series study.** *Dental Materials*, 32(8), 1042-1051.
- Lang, N. P., Berglundh, T., & Working Group 4 of the Seventh European Workshop on Periodontology. (2011). **Periimplant diseases: where are we now?—Consensus of the Seventh European Workshop on Periodontology.** *Journal of clinical periodontology*, 38, 178-181.
- Lopes, J. M. F. (2022). **Análise Elementar em Amostras de Saliva de Pacientes com Peri-Implantite por TXRF** (Master's thesis, Londrina State University).
- Mathew, M. T., Barão, V. A., Yuan, J. C. C., Assunção, W. G., Sukotjo, C., & Wimmer, M. A. (2012). **What is the role of lipopolysaccharide on the tribocorrosive behavior of titanium?.** *Journal of the mechanical behavior of biomedical materials*, 8, 71-85.
- Mercan, S., Bölükbaşı, N., Bölükbaşı, M. K., Yayla, M., & Cengiz, S. (2013). **Titanium element level in peri-implant mucosa.** *Biotechnology & Biotechnological Equipment*, 27(4), 4002-4005.
- Mombelli, A., Hashim, D., & Cionca, N. (2018). **What is the impact of titanium particles and biocorrosion on implant survival and complications? A critical review.** *Clinical Oral Implants Research*, 29, 37-53.
- Poletto, A. C., Singi, P., Barri, R. M., Casanova, A. A., Garbelini, C. C. D., da Silva, C. C., & Venancio, E. J. (2021). **Relationship of levels of trace elements in saliva and dental caries in preschool children using total reflection X-ray fluorescence technique (TXRF)*.** *Journal of Trace Elements in Medicine and Biology*, 63, 126663.
- Safioti, L. M., Kotsakis, G. A., Pozhitkov, A. E., Chung, W. O., & Daubert, D. M. (2017). **Increased levels of dissolved titanium are associated with peri-implantitis—a cross-sectional study.** *Journal of periodontology*, 88(5), 436-442.
- Saini, M., Singh, Y., Arora, P., Arora, V., & Jain, K. (2015). **Implant biomaterials: A comprehensive review.** *World Journal of Clinical Cases: WJCC*, 3(1), 52.
- Schwarz, F., Derks, J., Monje, A., & Wang, H. L. (2018). **Peri-implantitis.** *Journal of clinical periodontology*, 45, S246-S266.
- Soler, M. D., Hsu, S. M., Fares, C., Ren, F., Jenkins, R. J., Gonzaga, L., ... & Esquivel-Upshaw, J. F. (2020). **Titanium Corrosion in Peri-Implantitis.** *Materials*, 13(23), 5488.
- Van Grieken, R., & Markowicz, A. (Eds.). (2001). *Handbook of X-ray Spectrometry*. CRC press.
- Zahir, S., & Sarkar, S. (2006). **Study of trace elements in mixed saliva of caries free and caries active children.** *Journal of Indian Society of Pedodontics and Preventive Dentistry*, 24(1), 27.